A probabilistic modeling approach to assess human inhalation exposure risks to airborne aflatoxin B$_1$ (AFB$_1$)

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Abstract

To assess how the human lung exposure to airborne aflatoxin B$_1$ (AFB$_1$) during on-farm activities including swine feeding, storage bin cleaning, corn harvest, and grain elevator loading/unloading, we present a probabilistic risk model, appraised with empirical data. The model integrates probabilistic exposure profiles from a compartmental lung model with the reconstructed dose–response relationships based on an empirical three-parameter Hill equation model, describing AFB$_1$ cytotoxicity for inhibition response in human bronchial epithelial cells, to quantitatively estimate the inhalation exposure risks. The risk assessment results implicate that exposure to airborne AFB$_1$ may pose no significance to corn harvest and grain elevator loading/unloading activities, yet a relatively high risk for swine feeding and storage bin cleaning. Applying a joint probability function method based on exceedence profiles, we estimate that a potential high risk for the bronchial region (inhibition $= 56.69\%$ with 95% confidence interval (CI): 35.05–72.87%) and bronchiolar region (inhibition $= 44.93\%$ with 95% CI: 21.61 – 66.78%) is alarming during swine feeding activity. We parameterized the proposed predictive model that should encourage a risk-management framework for discussion of carcinogenic risk in occupational settings where inhalation of AFB$_1$-contaminated dust occurs.

Keywords: Aflatoxins; Mycotoxins; Risk assessment; Lung; Probabilistic

1. Introduction

Large gaps remain in the knowledge base needed to conduct quantitative risk assessment for inhaled mycotoxins (Wu, 2004). Case reports and studies of agricultural workers indicate that certain health effects occur from inhalation of molds that are due at least in part to mycotoxins. There are several case reports and epidemiological articles in which toxin-producing molds have been reported to be associated with health effects in indoor environments. The fungus *Aspergillus flavus* mainly produces aflatoxins. Aflatoxin B$_1$ (AFB$_1$), the most toxic of the aflatoxins, a mycotoxin contaminant commonly found in a variety of foods and feeds, is immunotoxic and carcinogenic in many animal models and is strongly suspected to be a human carcinogen (Bondy and Pestka, 2000). Although AFB$_1$ is a well-studied mycotoxin, exposures to AFB$_1$-containing dust have not been reported in indoor environments, and it is not known whether exposure to AFB$_1$ poses a health risk in indoor environments.

High concentrations of the carcinogen AFB$_1$ are commonly found in respirable, airborne dust, and inhaled AFB$_1$ has been shown to be a risk factor for...
occupational pulmonary carcinogenesis. There is some epidemiological evidence linking pulmonary exposure to AFB1-laden grain dust with an increase in lung tumor incidence in certain occupational settings (Desai and Ghosh, 2003; Ghosh et al., 1997). Kelley et al. (1997) indicated that in situ AFB1 activation and resultant carcinogenic risk are distinctly possible in occupational settings where inhalation of AFB1-contaminated dust occurs. The fate of AFB1 exposure via the respiratory tract is therefore of interest in an evaluation of potential occupational risk. Coulombe et al. (1991) have used a pharmacokinetic model to determine the disposition of AFB1 bound to respirable grain dust, suggesting that particle association of AFB1 increased the respiratory tract retention of this compound at early time intervals, which might be a factor in the reputed carcinogenic action of this compound in the respiratory tract.

Some evidence suggests that the human lung may be a target tissue for the action of AFB1 (Kelly et al., 1997). Two studies indicated that workers at a peanut- and linseed-processing plant, who were exposed to 0.04–2.5 μg of airborne AFB1 per 45-h week, experienced a higher incidence of upper respiratory (trachea and bronchus) tumors compared to unexposed cohorts (Van Nieuwenhuiize et al., 1973). A more comprehensive retrospective study showed no excess of respiratory cancer in workers at livestock feed processing plants exposed to an estimated 170 ng airborne AFB1 per day (Olsen et al., 1988). Agricultural surveys show that AFB1 in dust particles from grain mills can reach concentrations as high as 4708 ppb (McMillian et al., 1978). Predicted occupational exposure in a corn processing plant containing 107 ng of AFB1 m⁻³, and the daily occupational exposure to AFB1 was calculated to be from 40–856 ng, based on a respiration rate of 1 m³/h (Burg et al., 1981, 1982). Selim et al. (1998) indicated that airborne AFB1 found in dust collected during harvest and grain loading/unloading ranged from 0.04 to 92 ng m⁻³ and higher levels of AFB1 were found in the airborne dust samples collected from enclosed animal feeding buildings (5–421 ng m⁻³) and during bin cleaning (124–4849 ng m⁻³). Selim et al. (1998) suggested that farmers and farm workers might be exposed to potentially hazardous concentrations of AFB1, particularly during bin cleaning and animal feeding in enclosed buildings.

The objectives of this study are twofold: (1) to conduct an environmental risk assessment based on the USEPA methodology, and (2) to address the uncertainties by using a probabilistic approach to risk characterization that yields quantitative estimates of the risks themselves and also of their associated uncertainties. We reanalyze published data of airborne AFB1 measurements during selected on-farm activities and incorporate a compartmental lung model to estimate the AFB1 concentrations in lung cells. We combine predicted lung cell concentrations and a dose–response relationship derived from published experimental studies on human lung cells allowing us to assess risk endpoint. To determine overall uncertainty in predicted risks, the uncertainties resulting from the assessments of exposure and dose–response are propagated through the risk characterization process using Monte Carlo (MC) analysis.

2. Material and methods

Our probabilistic risk assessment framework is divided into four phases (Fig. 1) and is described in the subsequent sections.

2.1. Problem formulation: data reanalysis

The occupational settings focus on four selected on-farm activities including indoor: swine feeding and storage bin cleaning and outdoor: corn harvest and grain elevator loading/unloading. The major database is adopted from Burg et al. (1982) and Selim et al. (1998). The size distributions of airborne Aspergillus spp. in indoor and outdoor activities are reanalyzed and optimal fitted to the published data adopted from Gorny et al. (1999) and Sanchez-Monedero and Stentiford (2003). AFB1 concentration distributions of indoor/outdoor on-farm activities also determined followed the fitted size distributions along with the reported concentration data. We use Kolmogorov–Smirnov (K–S) statistics to optimize the goodness-of-fit of distribution of observed data by using the Statistica® software package (StatSoft, Tulsa, OK, USA).

2.2. Exposure analysis

We use a compartmental lung model to estimate AFB1 concentration in lung tissue (Liao et al., 2003). We divided the human respiratory tract (HRT) into five major compartments from the suggestion of ICRP66 (ICRP, 1994): (i) the nasal passage (ET1), comprising the anterior nose and the posterior nasal passages; (ii) the pharynx (ET2), comprising larynx and mouth; (iii) the bronchial region (BB), comprising the airway from the trachea, main bronchi, and intrapulmonary bronchi; (iv) the bronchiolar region (bb), comprising the bronchioles and terminal bronchioles; and (v) the alveolar-interstitial region (AI), comprising the airway from respiratory bronchioi through alveolar sacs. Followed by the principle of mass balance, the dynamic equations of inspiratory oral cavity (IOC) varying with particle size range k and time t to each regional compartment are given a by a linear dynamic equation (Liao et al., 2003; Chen et al., 2004). We solve the linear dynamic equation explicitly as AFB1 concentrations...
reach steady-state and yield the steady state AFB₁ concentration in each compartment as

\[ C(k) = -L(k)^{-1}B[C_1(k)]. \]  

(1)

where \( C(k) = \{C_1(k) C_2(k) C_3(k) C_4(k) C_5(k)\}^T \) is the state variable vector of AFB₁ concentrations in compartments ET₁, BB, bb, and AI, respectively (ng m⁻³); \( C_1(k) \) is the input AFB₁ concentrations (ng m⁻³); the constant

Fig. 1. Flowchart of probabilistic risk assessment framework to assess human inhalation exposure risk for AFB₁. The meanings of the symbols are described in the text.
input matrix \( [B] = \text{diag}(Q/V_1, 0, 0, 0) \); and the state matrix \( [L(k)] \) has the form as

\[
\begin{pmatrix}
-\lambda_{d}(k) - \lambda_{a}(k) - \lambda_{im}(k) \\
-\varepsilon_{1}(k) \frac{\partial}{\partial t} - \beta_{31} \frac{\partial}{\partial t} - \frac{\partial}{\partial t} \\
\beta_{31} \frac{\partial}{\partial t} \\
0 \\
0
\end{pmatrix}
\begin{pmatrix}
\frac{\partial}{\partial t} \\
\beta_{31} \frac{\partial}{\partial t} \\
\beta_{43} \frac{\partial}{\partial t} \\
\beta_{54} \frac{\partial}{\partial t} \\
\beta_{54} \frac{\partial}{\partial t}
\end{pmatrix}
\begin{pmatrix}
0 \\
\beta_{31} \frac{\partial}{\partial t} \\
\beta_{43} \frac{\partial}{\partial t} \\
\beta_{54} \frac{\partial}{\partial t} \\
-\lambda_{d}(k) - \lambda_{a}(k) - \lambda_{im}(k)
\end{pmatrix}
\begin{pmatrix}
0 \\
0 \\
\beta_{43} \frac{\partial}{\partial t} \\
\beta_{54} \frac{\partial}{\partial t} \\
-\lambda_{d}(k) - \lambda_{a}(k) - \lambda_{im}(k)
\end{pmatrix}
\]

in that \( Q \) is the breathing rate (cm\(^3\) h\(^{-1}\)); \( V_i \) is the volume of compartment \( i \) (cm\(^3\)); \( \beta_{in} \) is the transition coefficient from compartments \( n \) to \( m \); \( \lambda_{d}(k), \lambda_{a}(k), \) and \( \lambda_{im}(k) \) represent turbulent diffusive deposition rate, gravitational settling rate, and inertial impaction rate, respectively, in the \( k \)th size range in the compartment \( i \) (s\(^{-1}\)); \( \varepsilon_{1}(k) \) is the interception deposition efficiency in the \( k \)th size range in the compartment \( i \); and \( \lambda_{L}(t) \) is the time-dependent fungal spores clearance rate in the compartment \( AI \) (s\(^{-1}\)).

The major route of entry into the body of airborne AFB\(_1\) in the on-farm activities is inhalation, and this causes deposition and accumulation in HRT. We employ turbulent diffusive deposition rate equations of particulate AFB\(_1\). Inspiratory/expiratory oral cavity (IOC/EOC) and inspiratory/expiratory nasal–pharyngeal (INP/ENP) were treated as the breathing patterns during on-farm activities. Other physiological parameters, including clearance rate, transfer coefficient between lung compartments, and airways reference values, are obtained from ICRP66 (ICRP, 1994).

Table 1 summarizes the lung physiological parameters and the durations, frequencies and respiratory rates for four different on-farm activities used in the present analysis. We considered that storage bin cleaning, corn harvest, and grain elevator loading/unloading are heavy exercises and swine feeding is a light exercise (Table 1).

<table>
<thead>
<tr>
<th>Lung physiological parameter( ^a )</th>
<th>Description</th>
<th>Representation values</th>
</tr>
</thead>
<tbody>
<tr>
<td>( Q ) ( _t )</td>
<td>Breathing frequency</td>
<td>15, 20 breaths min(^{-1})</td>
</tr>
<tr>
<td>( V_i )</td>
<td>Tidal volume</td>
<td>1.33, 3 L</td>
</tr>
<tr>
<td>( C_L )</td>
<td>Clearance rate by phagocyte</td>
<td>( 8.3 \times 10^{-3} \text{h}^{-1} )</td>
</tr>
<tr>
<td>( \beta_{ij} )</td>
<td>Transfer coefficient between compartments ( i ) and ( j )</td>
<td>0.9–1.1</td>
</tr>
<tr>
<td>( D_1, D_2, D_3, D_4, D_5 )</td>
<td>Diameter of airways</td>
<td>0.5, 2.3, 1.2, 0.1, 0.05 cm</td>
</tr>
<tr>
<td>( n_1, n_2, n_3, n_4, n_5 )</td>
<td>Number of airways</td>
<td>1, 1, 1, ( 6.5 \times 10^5 ), ( 4.5 \times 10^7 )</td>
</tr>
<tr>
<td>( V_1, V_2, V_3, V_4, V_5 )</td>
<td>Volume of compartments in lung</td>
<td>5.8, 82.1, 94.6, 510.2, 1580.4 cm(^3)</td>
</tr>
<tr>
<td>On-farm activity</td>
<td>Duration (h ( \text{d}^{-1} ))</td>
<td>Frequency (d ( \text{yr}^{-1} ))</td>
</tr>
<tr>
<td>Swine feeding</td>
<td>2( ^b )</td>
<td>365( ^b )</td>
</tr>
<tr>
<td>Storage bin cleaning</td>
<td>4/bin( ^b )</td>
<td>2 bins( ^b )</td>
</tr>
<tr>
<td>Corn harvest</td>
<td>12( ^b )</td>
<td>7( ^b )</td>
</tr>
<tr>
<td>Grain elevator loading/unloading</td>
<td>12( ^c )</td>
<td>7( ^c )</td>
</tr>
</tbody>
</table>

\( ^a \)Adapted from ICRP66 (ICRP, 1994).
\( ^b \)Adapted from Selim et al. (1998).
\( ^c \)Estimated from Selim et al. (1998).
2.3. Effect analysis

Inhibition response of human lung cells in relation to cytotoxicity of AFB1 at low doses for the bronchial epithelial cell type is adapted from the published literature in that the cytochrome P-450 (CYP) 1A2-expressing human lung cells (B-CMV1A2) was the most susceptible cell type to the cytotoxic effects of AFB1 (Van Vleet et al., 2001, 2002). They concluded that human lung cells expressing these CYP isozymes are capable of activating AFB1, even at low environmentally relevant concentrations. They also suggested that any assessment of risk posed by inhaled AFB1 should take into account the relative expression of these isozymes in the human lung and it is possible that inhalation of AFB1 may result in an increased risk of lung cancer in exposed persons.

Van Vleet et al. (2002) used an empirical three-parameter Hill equation model to represent the cytotoxicity plots of % inhibition versus μM AFB1 (1 ng m⁻³ of AFB1 is equal to 3.2 × 10⁻⁶ μM m⁻³ based on the molecular weight of AFB1 = 312.3 g M⁻¹)

\[
I = \frac{I_{\text{max}} \times [\text{AFB1}]^{0.774}}{[\text{IC50}^{0.774} + [\text{AFB1}]^{0.774}} \quad (r^2 = 0.998), \tag{3}
\]

where \(I\) is the measured response (% inhibition), IC50 is the AFB1 concentration yielding half of the maximal response (\(I_{\text{max}} = 83.344\%\) inhibition) in that IC50 = 0.065 ± 0.02 (mean ± SD), 0.774 is the Hill coefficient \(n\) which is a measure of cooperativity in which an \(n < 1\) represents a supralinear response. We treated IC50 in Eq. (3) probabilistically to account for the inherent uncertainty that arises from a number of sources, including the limited number of observations and limited sample size within treatment sets.

To account for this uncertainty, we construct a distribution for the input variable of IC50. We determine normal distribution for IC50 and incorporate the distribution into the Monte Carlo (MC) simulation to obtain 2.5th and 97.5th percentiles as the 95% confidence interval (CI) for reconstructed dose-response profile. Uncertainty and/or variability were not considered for the reported Hill coefficient. This was unfortunate but unavoidable since the Hill coefficient from the published study was reported only as an average value. As a result, the risk curves and CI reported here do not incorporate this source of uncertainty. Applying the Hill equation model, the cumulative distribution function (cdf) of predicted cytotoxicity (% inhibition) function for a given AFB1 dose in human lung cell \(I(D)\), \(F(I|D)\), could be expressed symbolically as a conditional cdf:

\[
F(I|D) = \Phi\left(\frac{83.344 \times D^{0.774}}{(\text{IC50})^{0.774} + D^{0.774}}\right), \tag{4}
\]

where \(\Phi(\cdot)\) is the cumulative standard normal distribution.

2.4. Risk characterization

Risk characterization is the phase of risk assessment where the results of the exposure and quantitative effects assessments are integrated to provide an estimate of risk for the population under study. In this case, it entails combining the exposures, measured as AFB1 dose in human lung cell, with the quantitative dose-response relationship between lung cell AFB1 dose and associated % inhibition determined from the experimental studies.

Risk at a specific AFB1 dose in the lung cell \(D\) can be calculated as the proportion of the lung cell expected to have that cell concentration multiplied by the conditional probability of % inhibition, at a given dose, \(D\). This results in a joint probability function (JPF) or exceedence profile, which describes the probability of exceeding the concentration associated with a particular degree of effect. Graphic display of the JPF also provides a means of assessing how alterations in ambient concentrations due to management efforts would affect the risk assessment. This can be expressed mathematically as a probabilistic risk model as

\[
R(D) = F(D) \times F(I|D), \tag{5}
\]

where \(R(D)\) is the risk at a specific AFB1 dose \(D\), \(F(D)\) is the cdf of having lung cell AFB1 dose, and \(F(I|D)\) is the

![Fig. 2. Box and whisker plots of AFB1 concentration in (A) indoor on-farm activities of storage bin cleaning and swine feeding and in (B) outdoor on-farm activities of grain elevator loading/unloading and corn harvest.](image-url)
conditional cdf of the % inhibition, given lung cell AFB₁ dose $D$. A risk diagram was generated from the cumulative distribution of simulation outcomes. Each point on the risk diagram represents both the probability that the chosen proportion of lung cell will be affected and also the frequency with which that level of effect would be exceeded.

3. Results and discussion

3.1. Exposure analysis

Fig. 2 shows the box plots of interquartile and 50th percentile predictions associated with whisker plots indicating 5th- and 95th-percentile predictions of AFB₁ levels in on-farm indoor (storage bin cleaning and swine feeding) and outdoor (grain elevator loading/unloading and corn harvest) activities in that the particle size distributions are shown in Fig. 3. Fig. 3 indicates that particle size distributions of $A.\ flavus$ for on-farm indoor activities of swine feeding and storage bin cleaning have a lognormal distribution with a geometric mean diameter of 2.81 $\mu m$ and a geometric standard deviation of 1.65. Gorny et al. (1999) and Sanchez–Monedero and Stentiford (2003) reported that the major AFB₁-induced fungal spores of $A.\ flavus$ had their maximum concentrations in the aerodynamic size range 2.1–3.3 $\mu m$.

The distributions of AFB₁ level during storage bin cleaning, swine feeding, and corn harvest were more highly skewed at higher concentrations (Fig. 2), indicating that measured AFB₁ concentrations had a higher uncertainty as quantified by the variances in that the 5th- and 95th-percentiles predictions for storage bin cleaning and swine feeding are ranged from 225.31 to 3867.65 and 23.97 to 518.79 ng m⁻³, respectively, during
indoor on-farm activities; whereas during outdoor on-farm activities ranged from 9.74 to 23.21 and 0.07 to 1.59 ng m\(^{-3}\) for grain elevator loading/unloading and corn harvest, respectively. Fig. 2 also demonstrates that the magnitudes of measured median AFB\(_1\) concentrations during indoor activities of storage bin cleaning (10\(^3\)) is one order higher than that during swine feeding (10\(^2\)), whereas during outdoor activities, the AFB\(_1\) level of grain elevator loading/unloading (10\(^1\)) is two orders higher than that of corn harvest (10\(^{-1}\)). The differences of AFB\(_1\) level at four settings may be due to the existing environmental effects of different temperature and humidity. The higher the humidity, the higher the fungal growth of Aspergillus spores, especially for uncontrolled humidity condition in storage bins.

3.2. Dose–response model for human lung cells

The reconstructed dose–response profile (Fig. 4) was implemented by 5000 iterations of a MC simulation providing an adequate fit for the data points of AFB\(_1\) concentrations from 0 to 1 \(\mu\)M (\(\chi^2\) goodness of fit, \(P > 0.5\)). It can be seen from Fig. 4 that the calculated inhibition concentration inducing 50% inhibition (IC50) value is 0.065 \(\mu\)M with a 95% CI of 0.03 to 0.11 \(\mu\)M from the fitted dose–response model.

At present the dose–response relationships for fungal particles are understood poorly, i.e., the number of particles of each fungal genus or species needed to cause a certain symptom or disease is not known. Therefore, the suitability of published data for dose–response modeling still needs to be justified.

3.3. Risk estimates for respiratory deposition

Figs. 5A, C, and E show histograms for the predicted pdfs of AFB\(_1\) concentrations in different HRT regions of BB, bb, and AI during different indoor and outdoor on-farm activities. The relative skewness and spread in modeled output varied with on-farm activities. A box and whisker plot represents the uncertainty in comparing % inhibition for different on-farm activities in different HRT regions (Figs. 5B, D, and F). We first reanalyze published data of airborne AFB\(_1\) measurements of selected on-farm activities and then incorporate a compartmental lung model to estimate the AFB\(_1\) concentrations in lung cells. We solve the linear dynamic equation explicitly as AFB\(_1\) concentrations reach steady state and yield the steady-state AFB\(_1\) concentration in each compartment as shown in Eq. (1). We thus employ the MC simulation to predict the pdfs value of \(\{C_f(k)\}\) based on the input parameter of \(\{C(k)\}\) featuring a lognormal distribution.

We calculated the overall expected inhibition subjected to a mean AFB\(_1\) level to highlight the expected risk in different HRT regions during various on-farm

![Fig. 4. Reconstructed dose–response profile with 95% confidence interval optimally fitted by three-parameter Hill model equation in that AFB\(_1\) concentrations ranged from (A) 0 to 12 \(\mu\)M and (B) 0 to 1 \(\mu\)M.](image)
Fig. 5. Probabilistic density functions predicted for AFB$_1$ concentrations in different HRT regions: (A) BB, (C) bb, and (E) AI for four on-farm activities in that A1 = corn harvest, A2 = grain elevator loading/unloading, A3 = storage bin cleaning, and A4 = swine feeding. Box and whisker plots represent the uncertainty in comparing % inhibition for different on-farm activities in different HRT regions: (B) BB, (D) bb, and (F) AI.
Table 2
The overall expected inhibition (I (%)) subjected to a mean AFB1 level (μM) in different lung regions of BB, bb, and AI during four different on-farm activities

<table>
<thead>
<tr>
<th></th>
<th>Swine feeding</th>
<th>Storage bin cleaning</th>
<th>Corn harvest</th>
<th>Grain elevator loading/unloading</th>
</tr>
</thead>
<tbody>
<tr>
<td>BB</td>
<td>AFB1</td>
<td>I</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>1.59 × 10⁻¹</td>
<td>56.69 (35.05–72.87)⁹</td>
<td>2.92 × 10⁻²</td>
<td>1.09 × 10⁻⁴</td>
</tr>
<tr>
<td></td>
<td>2.92 × 10⁻²</td>
<td>30.37 (13.82–51.45)</td>
<td>1.09 × 10⁻⁴</td>
<td>4.90 × 10⁻³</td>
</tr>
<tr>
<td>BB</td>
<td>4.90 × 10⁻³</td>
<td>2.23 (4.39–8.25)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>bb</td>
<td>AFB1</td>
<td>I</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>7.34 × 10⁻²</td>
<td>44.93 (21.61–66.78)</td>
<td>1.34 × 10⁻²</td>
<td>5.12 × 10⁻⁵</td>
</tr>
<tr>
<td></td>
<td>1.34 × 10⁻²</td>
<td>19.90 (8.18–39.79)</td>
<td>5.12 × 10⁻⁵</td>
<td>2.23 × 10⁻³</td>
</tr>
<tr>
<td>bb</td>
<td>2.23 (4.39–8.25)</td>
<td>6.04 (1.31–1.34)</td>
<td>6.04 (1.31–1.34)</td>
<td></td>
</tr>
<tr>
<td>AI</td>
<td>AFB1</td>
<td>I</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2.60 × 10⁻²</td>
<td>28.62 (10.78–52.55)</td>
<td>4.68 × 10⁻³</td>
<td>7.80 × 10⁻⁴</td>
</tr>
<tr>
<td></td>
<td>4.68 × 10⁻³</td>
<td>10.16 (3.67–24.21)</td>
<td>7.80 × 10⁻⁴</td>
<td>3.79 (2.02–3.84)</td>
</tr>
</tbody>
</table>

⁹95% CI is calculated from 2.5th and 97.5th-percentiles of 5000 MC simulations.

Fig. 6. Exceedence risk diagrams with 95% confidence interval of four on-farm activities of (A) corn harvest, (B) grain elevator loading/unloading, (C) swine feeding, and (D) storage bin cleaning in different HRT regions of BB, bb, and AI.
activities (Table 2). Table 2 suggests that the relatively high risk for regions BB (I = 56.69% with 95% CI: 35.05–72.87%) and bb (I = 44.93% with 95% CI: 21.61–66.78%) is alarming during swine feeding activity.

Risk curves shown in Fig. 6 indicate the estimated probabilistic of inhibitions of differing on-farming activities for different HRT regions. The plotted probabilities, calculated from the outcome of the MC simulation followed a JPF shown in Eq. (5) describing the exceedence cdfs (Fig. 6) associated with a dose–response relationship (Fig. 4), taking into account the uncertainty in estimating risk. Fig. 6 demonstrates that the probabilities that 10% or more of the lung cells in regions BB, bb, and AI (risk = 0.1) affected during swine feeding activity are approximately 71%, 62%, and 48%, respectively, with 95% CI of 65–75%, 54–70%, and 38–58%, respectively. Generally, for corn harvest, grain elevator loading/unloading, and storage bin cleaning, the probability is 0.1 that at least 0.4–1.6%, 4–13%, and 21–48% inhibition, respectively, exist for lung cells in AI, bb, and BB regions.

We believe that a probabilistic risk-based framework—probability distributions and risk diagrams such as Fig. 6—is an effective representation of state-of-the-art results of scientific assessments for human response to airborne AFB1 exposure during on-farm activities. To our knowledge, this risk-based framework has not been addressed until now. Although the suitability and effectiveness of techniques for presenting uncertain results is context dependent, we believe that such probabilistic methods are more valuable for communicating an accurate view of current scientific knowledge to those seeking information for decision-making than assessments that do not attempt to present results in probabilistic framework. We suggest that our probabilistic framework and methods be taken seriously because they produce general conclusions that are more robust than estimates made with a limited set of scenarios or without probabilistic presentations of outcomes, and our modeling technique offers a risk-management framework for discussion of future establishment of limits for respiratory exposure to airborne AFB1.

References